

# **RESEARCH ARTICLE**

# A Case Series of Hypertrophic Cardiomyopathy Conducted in Vietnam Revealing a Novel Pathogenic Variant of the TNNT2 Gene

Hung Manh Pham<sup>1,2,#</sup>, Van Khanh Tran<sup>1,#</sup>, Trung Anh Mai<sup>2</sup>, Long Hoang Luong<sup>1,3</sup>, May Le Pham<sup>4,5</sup>, Chi Khanh Nguyen<sup>4,6</sup>, Hoai Thu Thi Nguyen<sup>2</sup>, Minh Nhat Pham<sup>2</sup>, Can Thuy, Do<sup>2</sup>, Thanh Tuan Le<sup>2</sup>, Thanh Van Ta<sup>1</sup> and Thinh Huy Tran<sup>1,\*</sup>

<sup>1</sup>Center for Gene and Protein Research, Hanoi Medical University, Hanoi, Vietnam

<sup>2</sup>Department of Cardiology, Vietnam National Heart Institute, Bach Mai Hospital, Hanoi, Vietnam

<sup>3</sup>Department of Biology, Hanoi-Amsterdam High School for the Gifted, Hanoi, Vietnam

<sup>4</sup>Vietnam National University, VNU University of Medicine and Pharmacy, Hanoi, Vietnam

<sup>s</sup>Department of of Health Sciences, Vin University, Hanoi, Vietnam

<sup>6</sup>Department of Allergy & Clinical Immunology, E Hospital, Hanoi, Vietnam

# Abstract:

# Background:

Hypertrophic Cardiomyopathy (HCM) is one of the leading causes of sudden cardiac death in adults. HCM is inherited in an autosomal dominant manner; however, the genetic etiology of the disease is not fully explained and studies on the hereditary characteristics in family trees are still underway.

## Methods:

Ten HCM patients and 31 of their relatives were recruited. Targeted sequencing for 4 HCM related-genes, including *MYH7*, *MYBPC3*, *TNNT2*, and *TNNI3*, using targeted next-generation sequencing (NGS) was carried out. Demographic, clinical, electrocardiography, and echocardiography characteristics were also characterized.

## Results:

Among the 10 HCM patients, 5 were identified with the HCM pathogenic variants in MYH7 (3 patients), MYBPC3 (1 patient), and TNNT2 (1 patient) genes. Eleven out of 31 relatives from these 5 genotype-positive patients carried the same pathogenic variants. We found the novel c.822-2 A>G variant in the splicing site of the *TNNT2* gene responsible for HCM disease in a family with 7 subjects genotype positive and 3 others who suffered from sudden cardiac death.

## Conclusion:

This case series highlighted the importance of genetic testing for clinically confirmed HCM patients and family members. The genetic information can be used as a molecular marker to complement the clinical presentation in the diagnosis of HCM, as well as a prognostic tool for the patients and their family members.

Keywords: Hypertrophic cardiomyopathy, Vietnamese, MYH7, MYBPC3, TNNT2, TNNT3, Genetic testing, Novel pathogenic variant.

| Article History Received: July 1, 2021 Revised: Nove | ber 10, 2021 Accepted: January 12, 2022 |
|--|---|
|--|---|

# **1. INTRODUCTION**

Hypertrophic cardiomyopathy (HCM) is recognized as the most common monogenic cardiovascular disease with a preva-

\* Address correspondence to this author at the Center for Gene and Protein Research, Hanoi Medical University, 1<sup>st</sup> Ton That Tung Street, Hanoi 10000, Vietnam; Tel: + 84 38523798/ 244; Fax: + 84 4 38525115; E-mail: tranhuythinh@hmu.edu.vn

*<sup>#</sup>These authors contributed equally to this work* 

lence of 1:500 in young adults. It is also one of the leading causes of sudden cardiac death in young adults [1, 2]. HCM is defined as the thickening of the myocardial wall in any segments of the left ventricle (LV) in the absence of any other causes [3]. HCM is inherited as an autosomal dominant genetic trait that could be transmitted to offspring with a proportion up to 50%. Recent studies have indicated that 60% of HCM patients could be identified with the sarcomere disease-causing

#### variant [4, 5].

The genetic etiology of HCM is not fully explained and studies on the genotype-phenotype correlation are still underway. Up until recently, 11 genes have been identified and extensively studied for their pathogenicity in HCM. Among them, pathogenic variants in a  $\beta$ -myosin heavy chain (*MYH7*) and cardiac myosin-binding protein C (*MYBPC3*) genes are the most commonly associated with HCM, followed by Cardiac Troponin T (*TNNT2*) and Cardiac Troponin I (*TNNT3*) genes [6, 7].

Recent existing data have been mostly obtained in Caucasian populations while there is a dearth of studies conducted on the Vietnamese population, particularly the population in the North of Vietnam. The study of Thu et al. focused on 23 disease-related genes has demonstrated a prevalence of HCM-related gene variants in Vietnamese people [8]. However, the hereditary characteristics in families as well as the clinical characteristics of probands and their relatives have not yet been clarified. Therefore, we carried out this study on a series of patients and family members, with an extended Next Generation Sequencing panel covering the 4 most prevalent HCM-related genes (MYH7, MYBPC3, TNNT2, and TNNI3). We also discussed the extent of penetrance and phenotype-genotype correlation of the variants found in our study and performed a literature review to assess differences across populations.

## 2. METHODS

This study was conducted at the Vietnam National Heart Institute (VNHI), Bach Mai hospital, Hanoi, Vietnam, from July 2018 to August 2019. Ten patients diagnosed clinically with HCM were recruited together with 31 family members. The study protocol was approved by the Ethics Committee of Hanoi Medical University (Hanoi Medical University Institutional Review Board, Hanoi, Vietnam, reference number: IRB00003121). The study was abided by the Declaration of Helsinki in regards to the study involving human subjects. Informed consent was obtained from all participants and parents (for patients under 18 years old).

## 2.1. Clinical Assessment

Patients were clinically diagnosed with HCM by experienced cardiologists and echo-cardiologists at the Vietnam NationalHeart Institute (VNHI). The diagnosis criteria were based on the 2014 (European Society of Cardiology) ESC guidelines defining it as the wall thickness of any left ventricle (LV) myocardial segments  $\geq 15$ mm with no other causes explaining the hypertrophy. For children, the diagnosis of HCM was based on an LV wall thickness of 2 standard deviations greater than the predicted mean (z-score>2) [9, 10]. Other clinical data, including symptoms, medical history, and family history, were also recorded. A 12-lead resting electrocardiogram was performed on all subjects (Nihon Kohden Model 1250A Cardiofax S, Nihon Kohden, Tokyo, Japan). The presence of atrial fibrillation (AF), LV hypertrophy, left or right bundle-branch block, left atrial, and/or ventricular enlargement, abnormal Q waves, and nonspecific ST-T segments were recorded. Patients with suspected paroxysmal arrhythmias (paroxysmal atrial fibrillation (PAF),

ventricular tachycardia, premature ventricular contractions (PVC), or atrioventricular block) were confirmed with ambulatory electrocardiogram monitored for 24 h. Echocardiography evaluation for patients and relatives was performed using GE Vivid E9 Ultrasound System (GE Healthcare, Boston, US) and EchoPAC Clinical Workstation Software version V202 (GE Healthcare, Boston, US). All the indexes, including measurements of LV dimensions and wall thickness (septal, posterior, and maximal left ventricular thickness), LV ejection fraction, right ventricular hypertrophy and right ventricular dynamic obstruction, LV diastolic function, left atrial volume index, pulmonary artery systolic pressure, dynamic obstruction at rest and combined with the Valsalva maneuver, mitral valve, and papillary muscle functions were evaluated according to recommendations of the 2011 American Society of Echocardiography [11].

### 2.2. Genetic Analysis

Upon enrollment, blood samples were obtained from each subject and collected into sterile EDTA (ethylenediamine tetraacetic acid) tubes for genotyping and laboratory analysis. Genomic DNA was isolated from a 2ml peripheral blood sample obtained from each subject using the Wizard® Genomic DNA Purification Kit (Promega, Madison, USA). DNA was stored at -80°C until the time of analysis. Samples were sent for Next-Generation Sequencing (NGS) of a small panel of 4 genes, including MYH7, MYBPC3, TNNT2, and TNNI3. Sanger's sequencing was the method of choice for verification of identified variants and testing of family members (Primers' sequence for Sanger's sequencing is available upon request). PCR amplification product (100-150 ng starting DNA) was obtained for each sample. After agarose gel discrimination, the PCR product was purified with Gel Purification Kit followed by sequencing using Big Dye Terminator V3.1 (Applied Biosystems, Massachusetts, US). Results were analyzed by CLC Main Workbench Software (Qiagen, Hilden, Germany).

#### 2.3. Variant Analysis

Genetics variants were evaluated for their pathogenicity based on the guideline of the American College of Medical Genetics [12]. Appropriate *in silico* tools and public domain libraries (ClinVar – Clinically relevant Variant, HGMD – Human Gene Mutation Database; LOVD - Leiden Open Variation Database) were utilized to confirm whether variants were previously reported [12]. Pymol was utilized to visualize the affected protein (as well as the location of the pathogenic variants (comparing the native and mutant amino acids) [13].

## **3. RESULTS**

Clinical characteristics, ECG, and echocardiography findings of 16 subjects having genetic positive results are shown in Table 1. We identified four reported pathogenic variants and a new variant of *TNNT2* in 5 families. The age of diagnosis for probands with confirmed pathogenic variants was  $29 \pm 14.7$  years for 5 patients with HCM-related pathogenic variants and  $64.8 \pm 4.5$  years for the group without any suspected pathogenic variant; the difference was significant (p=0.02). Other characteristics of the two groups of patients are shown in Table 2.

| S.No. | Mutation                         | Relation-<br>ship | Age | Sex | Symptoms                           | Cardiac<br>Phenotype                                    | ECG                                   | LVWT<br>(mm) | EF (%) | LVOT,<br>mmHg | Manageme<br>nt   |
|-------|----------------------------------|-------------------|-----|-----|------------------------------------|---|---------------------------------------|--------------|--------|---------------|--|
| 1     | p.T1377M<br><i>MYH7</i>          | Proband           | 62  | F   | Dyspnea,<br>syncope                | Septal<br>hypokinesis<br>(after ASA)                    | AF, LBBB                              | 20           | 58     | 60            | BBs and<br>ASA   |
|       |                                  | Son               | 40  | М   | DOE, chest<br>discomfort           | Sigmoid<br>septum with<br>no<br>obstruction             | NSR,<br>NSST-T                        | 18           | 73     | 5             | BBs  |
|       |                                  | Grandson          | 11  | М   | None                               | Normal  | NSR                                   | 8            | 82     | 9             | None   |
| 2     | p.R782H<br><i>MYH7</i>           | Proband           | 39  | М   | Dyspnea,<br>oedema,<br>syncope     | Septal<br>hypokinesis<br>(after ASA)                    | BAV III<br>PMR<br>VT in<br>Holter 24h | 22           | 37     | 55            | BBs, ASA,<br>and ICD<br>implantation                         |
|       |                                  | Brother           | 27  | М   | DOE, chest<br>discomfort           | Septum and<br>apex<br>hypertrophy,<br>no<br>obstruction | NSR,<br>NSST-T                        | 19           | 61     | 8.5           | BBs  |
| 3     | c.822-2A><br>G<br><i>TNNT2</i> * | Proband           | 25  | F   | DOE, chest<br>pain                 | Both<br>ventricular<br>hypertrophy<br>with LVOT         | RBBB                                  | 24           | 58     | 110           | Medical<br>treatment for<br>heart failure,<br>BBs and<br>ASA |
|       |                                  | Father            | 54  | F   | DOE,<br>syncope                    | RV<br>hypertrophy                                       | NSR<br>RVH                            | 15           | 65     | None          | Medical<br>treatment for<br>heart failure                    |
|       |                                  | Aunt              | 52  | М   | DOE                                | RV<br>hypertrophy                                       | BAV III                               | 14           | 68     | None          | Pace-maker<br>implantation<br>, BBs                          |
|       |                                  | Aunt              | 37  | F   | DOE                                | Septal and<br>RV<br>hypertrophy                         | NSR<br>NSST-T                         | 13.4         | 66     | 10            | BBs  |
|       |                                  | Uncle             | 41  | М   | None                               | Normal  | NSR,<br>NSST-T                        | 7            | 80     | None          | None   |
|       |                                  | Cousin            | 3   | F   | None                               | Normal  | Normal                                | 5            | 71     | None          | None   |
|       |                                  | Cousin            | 1   | М   | None                               | Normal  | Normal                                | 6            | 65     | None          | None   |
| 4     | p.D770N<br><i>MYBPC3</i>         | Proband           | 18  | F   | DOE,<br>palpitations               | Sigmoid<br>septum, no<br>obstruction                    | NSST-T<br>PAF in<br>Holter 24h        | 24           | 74     | 12            | BBs,<br>Acenocoum<br>arol                                    |
|       |                                  | Father            | 52  | М   | None                               | Normal  | NSR<br>RBBB                           | 10           | 69     | None          | BBs  |
| 5     | p.R1114H<br><i>MYH7</i>          | Proband           | 30  | М   | Dyspnea,<br>chest pain,<br>Syncope | Symmetric<br>hypertrophy                                | NSTT-T                                | 31           | 80     | 50            | BBs and<br>ASA   |
|       |                                  | Father            | 53  | М   | DOE                                | Sigmoid<br>septum, no<br>obstruction                    | NSR,<br>RVH                           | 19           | 82     | None          | BBs  |

| Table 1. Demographic, clinical, ECG, and echocardiography characteristics of patients having a positive genotype and their |  |
|--|--|
| relatives.   |  |

DOE, Dyspnea on exertion; RV, Right ventricular; AF, Atrial fibrillation; PAF, Paroxysmal atrial fibrillation; LBBB, Left bundle branch block; RBBB, Right bundle branch block; NSR, Normal sinus rhythm; NSST-T, Non-specific ST-T; ASA, Alcohol septal ablation; LVOT, Left ventricular outflow tract; RVH, Right ventricular hypertrophy; BAV, Atrioventricular block; BBs, Beta-blocker agents; ICD, Implantable cardioverter-defibrillation; M, male; F, female.

Table 2. Clinical characteristics of the hypertrophic cardiomyopathy patient cohort.

| Proband Clinical Characteristic              | Confirmed Pathogenic Variant (n=5) | No Genetic Findings (n=5) | р    |
|--|------------------------------------|---------------------------|------|
| Gender, male/female                          | 2 males/3 females                  | 3 males/2 females         | ns   |
| Age at diagnosis, years                      | $29 \pm 14.7$                      | $64.8 \pm 4.5$            | 0.02 |
| Syncope/Presyncope                           | 3                                  | 0                         | ns   |
| Cardiac symptoms (NYHA $\geq 2/CCS \geq 2$ ) | 4                                  | 4                         | ns   |

#### 4 The Open Cardiovascular Medicine Journal, 2022, Volume 16

#### (Table 2) contd.....

| Proband Clinical Characteristic | Confirmed Pathogenic Variant (n=5) | No Genetic Findings (n=5) | р    |
|---------------------------------|------------------------------------|---------------------------|------|
| Family history of SCD           | 3                                  | 1                         | ns   |
| Family history of HCM           | 5                                  | 3                         | ns   |
| Maximal LVWT, mm                | $17.9 \pm 10$                      | $17.1 \pm 1.9$            | ns   |
| Severe hypertrophic >20 mm      | 4                                  | 3                         | ns   |
| LVOT pressure, mmHg             | $18.2 \pm 12.5$                    | 55.6 ± 48                 | 0.08 |
| LVOT ≥30 mmHg                   | 4                                  | 4                         | ns   |
| LVOT ≥50 mmHg                   | 4                                  | 2                         | ns   |
| SAM                             | 4                                  | 4                         | ns   |
| Heart failure, LVEF <40%        | 1                                  | 0                         | ns   |
| ECG abnormalities               | 5                                  | 4                         | ns   |
| Treatment                       |                                    |                           |      |
| Medications only                | 1                                  | 3                         |      |
| ASA/Septal myectomy             | 4                                  | 2                         | ns   |
| ICD                             | 1                                  | 0                         |      |

NYHA, New York Heart Association; CCS, Canadian Cardiovascular Society; SCD, Sudden cardiac death; HCM, Hypertrophic cardiomyopathy; LVWT, Left ventricular wall thickness; LVOT, Left ventricular outflow tract; SAM, Systolic anterior motion; LVEF, Left ventricular ejection fraction; ECG, Echocardiography; ASA, Alcohol septal ablation; ICD, Implantable Cardioverter-Defibrillator.



Fig. (1). Pedigree of the family with novel c.822-2A>G mutation in the TNNT2 gene.

## 3.1. Results of Genetic Analysis

By fully sequencing 4 common genes related to HCM (*MYH7, MYBPC3, TNNT2,* and *TNNI3*), we identified genetic variants in 5 families that can be attributed to HCM. The variants were p.T1377M, p.R783H, p.R1114H on *MYH7*, p.D770N on *MYBCP3*, and c.822-2A>G on *TNNT2*.

Reconfirmation using Sanger's sequencing was performed on the proband as well as the family members to identify that all 5 variants found were in the heterozygous state and exist in at least 2 members of each family. The variants found are outlined in Table 1; the primer for Sanger's sequencing is available from the authors upon reasonable request. Four out of 5 variants have been identified in patients with HCM (p.T1377M, p.R783H, p.R1114H on *MYH7*, and p.D770N on *MYBCP3*) and one newly identified variant c.822-2A>G on *TNNT2* was identified. However, we still performed an assessment based on the ACMG (American College of Medical Genetics) guidelines [12] to classify the pathogenicity of variants. We assessed four reported variants based on ACMG criteria matched with previous pathogenicity reports available on databases, such as Clinvar, HGMD, or LOVD. The full criteria and classification are included in the Supplementary Tables. Here, we identified a novel variant not previously reported in patients with HCM, c.822-2A>G on *TNNT2*.

## 3.1.1. Synopsis of the Novel TNNT2 Pathogenic Variant

Fig. (1) shows the pedigree of the family with the novel c.822-2A>G variant in the *TNNT2* gene. The proband (subject IV.4) was a 25-year-old woman who had dyspnea on exertion and chest pain. Echocardiography showed left ventricular hypertrophy involving the septum, anterior, anterolateral walls, and also the apex with severe LV outflow tract obstruction (pressure gradient, 110 mmHg). She was treated with alcohol septal ablation (ASA) and beta-blockers (BBs), and the symptoms were alleviated (Fig. **2**).

At the time of research, 11 of her relatives were screened for HCM. Three of them, including the proband's father (subject III.6) and 2 of her aunts (subject III.3 and subject III.5), were clinically confirmed to have HCM and also carried the same pathogenic variant as the proband. Meanwhile, her uncle (subject III.1) and his children (subjects IV.1, IV.2) who were also a carrier of this variant had no clear indication of HCM. The patient's mother (subject III.7), her sister (subject IV.5), two of her aunts (subjects III.8 and III.11), and her uncle (subject III.12) did not have any manifestations of HCM and were genetically negative as confirmed by using the Sanger sequencing method.

Three other relatives (subjects II.1, II.3, and III.10) were reported to have suffered sudden cardiac death (SCD) at the age of 40, 63, and 30 years, respectively. All of them were asymptomatic and did not have a history of any diseases prior to their death. Clinical characteristics of subjects carrying the c.822-2A>G variant in the TNNT2 gene are shown in Table 1.

#### 4. DISCUSSION

In this study, HCM-causative variants of 4 genes, including *MYH7, MYBPC3, TNNT2,* and *TNNI3,* which encoded for the beta-myosin heavy chain, myosin-binding protein C, cardiac troponin T, and cardiac troponin I, respectively, were found to be associated with HCM in our cohort. Among 41 individuals from the families of 10 patients, 5 out of 10 patients (50%) carried a single variant related to *MYH7, MYBPC3,* and *TNNT2* genes and 11 (35.5%) of their relatives had the same variants. A novel c.822-2A>G variant of the splicing receptor zone of *TNNT2* was found in a family with 7 individuals positive with HCM genetic testing but only 4 out of these 7 individuals had HCM phenotype.

The prevalence of attributable pathogenic variants in our cohort having familial HCM was lower than the reported

prevalence in the US and French cohorts, which were 54.2% and 60.6%, respectively [13, 14], but higher than other cohorts in Portugal, Taiwan, Finland, and Japan, which accounted for 28%, 34.2%, 38.2%, and 43.8% [15 - 18], respectively. Recently, a study conducted on Vietnamese HCM patients in the South of Vietnam found a panel of 23 HCM-related genes which pointed out that 43.4% of patients diagnosed with HCM carried variants mostly focused on *MYH7*, *TPM1*, and *TNNT2* genes, which were linked with more severe clinical manifestations [8]. However, the difference in prevalence might be due to the limited number of cases in our cohort as well as the difference in gene panel used.

Many previous studies involving patients with familial HCM in Caucasian and Asian populations found mostly variants of MYBPC3 and MYH7. Our study noted that in 2 families which carried pathogenic variants of the MYH7 gene (p.T1377M and p.R782H), sudden cardiac deaths of family members occurred more frequently. Fujita et al. [19] reported Japanese patients with HCM carrying variants in the TNNT2 gene that were associated with a risk of sudden cardiac death in youth. In our cohort, family number 3 were found to carry the novel variant c.822-2A>G in the TNNT2 gene, which might have caused the SCD of 3 family members. In this family, 6 family members were carrying the same c.822-2A>G variant along with the patient, but only 3 of them expressed the HCM phenotype, including the patient's father and her 2 aunts. Three other individuals were genotype positive - phenotype negative, but 2 of them (subjects IV.1, IV.2 in Fig. (1) were under the age of 5 years.

*TNNT2* gene has 16 exons, and the novel c.822-2A>G mutation occurs at the exon-16 splice acceptor site, thus affecting only the ultimate C-terminal part of the protein. We were able to find one study that modeled *in vivo* human TNNT2 intron 15 mutation that causes two different mutants splice products of TNNT2 [20, 21]. The location of their pathogenic variant was in close proximity to the variant in our study (c.822-2G>A); in their study, the resulting morphant splice product excludes human Exon 16, causing a disruption in the C-terminus of the zebrafish TNNT2 protein at the identical position to that mutated in humans, and is thought to be involved in the interaction with tropomyosin. The morphant ventricles exhibit restrictive physiology, and diminished contractility and defective tnnt2 splicing cause sarcomeric disarray in the embryonic heart, leading to HCM.

The genes we chose to investigate in our panel were more frequently reported to be associated with HCM, and each has a direct role in the heart muscle constitution and function. However, we suspect that there would be other potentially novel pathogenic genes or pathogenic variants in the noncoding region which could be identified with an expanded panel of Whole Exome or Whole Genome Sequencing. In this study, due to the number of cases and short observation time, we were unable to gauge the difference in disease severity of HCM in relation to each disease-causing gene. Future studies should also focus on prospective follow-up of the patient to assess the disease's natural history in relation to its pathogenic variants.



Fig. (2). Cardiac morphology evaluated by echocardiography of subjects having c.822-2A>G mutation in the TNNT2 gene. Subject IV.4 (proband) had septal hypertrophy in the mid-segment; however, the basal septum was normal (after ASA). Subjects III.1, IV.1, and IV.2 did not show cardiac hypertrophy on Echo at the time of the research. Subject III.3 (proband's aunt) had mid-septal and right ventricular hypertrophy without LVOT obstruction. Both III.5 (proband's aunt) and III.6 (proband's father) had right ventricular and also borderline septal hypertrophy without LVOT obstruction.

## CONCLUSION

This study was conducted on a group of 10 probands with HCM and their family members. A targeted Next-Generation Sequencing panel was used, and in-depth clinical profiling was carried out to study the prevalence of genetic defects and the genotype-phenotype correlation in this group of patients. This case series highlights the importance of genetic testing for clinically confirmed HCM patients and family members. The genetic information can be used as a molecular marker to complement the clinical presentation for the diagnosis of HCM, as well as a prognostic tool for the patients and their family members.

## **AUTHORS' CONTRIBUTION**

Hung Manh Pham, Van Khanh Tran, Trung Anh Mai, and Long Hoang Luong conceived and designed the study and performed the analysis. Thanh Van Ta and Huy Thinh Tran contributed to the development of the study's idea and design. Hoai Thu Nguyen Thi, May Le Pham, and Chi Khanh Nguyen contributed to data collection and carried out the experiments. Minh Nhat Pham, Can Thuy Do, and Thanh Tuan Le performed analysis and finalized the results. Hung Manh Pham, Van Khanh Tran, Long Hoang Luong, and Trung Anh Mai contributed to the drafting of the manuscript. All authors have read and approved the final version for publication.

# LIST OF ABBREVIATIONS

| ETHICS | ٨D | DOVAL AND CONSENT              |  |  |
|--------|----|--------------------------------|--|--|
| SCD    | =  | Sudden Cardiac Death           |  |  |
| ESC    | =  | European Society of Cardiology |  |  |
| ECG    | =  | Electrocardiography            |  |  |
| LV     | =  | Left Ventricular               |  |  |
| HCM    | =  | Hypertrophic Cardiomyopathy    |  |  |

# ETHICS APPROVAL AND CONSENT TO PARTI-CIPATE

The study protocol was approved by the Ethics Committee of Hanoi Medical University (Hanoi Medical University Institutional Review Board, Hanoi, Vietnam, reference number: IRB00003121).

## HUMAN AND ANIMAL RIGHTS

No animals were used that are the basis of this research. The study complies with the Declaration of Helsinki regarding the use of human samples and identifiable information.

## CONSENT FOR PUBLICATION

The patients gave consent to publish their information, including clinical and genetics information. No identifiable information was disclosed in any form.

#### AVAILABILITY OF DATA AND MATERIALS

The datasets used and/or analyzed during the current study are available from the corresponding author, [T.H.T] upon reasonable request. We, however, cannot provide personal information or data containing the identification of the patients in any form.

## FUNDING

None.

## CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

# ACKNOWLEDGEMENTS

The authors would like to sincerely thank the patients and their parents for allowing them to publish the data. They would also like to thank their colleagues who helped them in patients recruitment and clinical data collection.

## SUPPLEMENTARY MATERIAL

Supplementary material is available on the publisher's website along with the published article.

## REFERENCES

- [1] Maron BJ, Roberts WC, Epstein SE. Sudden death in hypertrophic cardiomyopathy: a profile of 78 patients. Circulation 1982; 65(7): 1388-94
- [http://dx.doi.org/10.1161/01.CIR.65.7.1388] [PMID: 7200406] [2] Maron BJ, Gardin JM, Flack JM, Gidding SS, Kurosaki TT, Bild DE. Prevalence of hypertrophic cardiomyopathy in a general population of young adults. Echocardiographic analysis of 4111 subjects in the CARDIA Study. Coronary artery risk development in (Young) adults. Circulation 1995: 92(4): 785-9 [http://dx.doi.org/10.1161/01.CIR.92.4.785] [PMID: 7641357]
- [3] Elliott P, Andersson B, Arbustini E, et al. Classification of the cardiomyopathies: a position statement from the European Society Of Cardiology Working Group on Myocardial and Pericardial Diseases. Eur Heart J 2008; 29(2): 270-6. [http://dx.doi.org/10.1093/eurheartj/ehm342] [PMID: 17916581]
- Richard P, Charron P, Carrier L, et al. Hypertrophic cardiomyopathy: [4] distribution of disease genes, spectrum of mutations, and implications for a molecular diagnosis strategy. Circulation 2003; 107(17): 2227-32

[http://dx.doi.org/10.1161/01.CIR.0000066323.15244.54] [PMID: 12707239]

- Van Driest SL, Ellsworth EG, Ommen SR, Tajik AJ, Gersh BJ, [5] Ackerman MJ. Prevalence and spectrum of thin filament mutations in an outpatient referral population with hypertrophic cardiomyopathy. Circulation 2003; 108(4): 445-51. [http://dx.doi.org/10.1161/01.CIR.0000080896.52003.DF] [PMID: 128609121
- [6] Bos JM, Towbin JA, Ackerman MJ. Diagnostic, prognostic, and therapeutic implications of genetic testing for hypertrophic cardiomyopathy. J Am Coll Cardiol 2009; 54(3): 201-11. [http://dx.doi.org/10.1016/j.jacc.2009.02.075] [PMID: 19589432]
- Maron BJ, Maron MS, Semsarian C. Genetics of hypertrophic [7] cardiomyopathy after 20 years: clinical perspectives. J Am Coll Cardiol 2012; 60(8): 705-15. [http://dx.doi.org/10.1016/j.jacc.2012.02.068] [PMID: 22796258]
- Tran Vu MT, Nguyen TV, Huynh NV, Nguyen Thai HT, Pham [8] Nguyen V, Ho Huynh TD. Presence of hypertrophic cardiomyopathy related gene mutations and clinical manifestations in vietnamese patients with hypertrophic cardiomyopathy. Circ J 2019; 83(9): 1908-16.

[http://dx.doi.org/10.1253/circj.CJ-19-0190] [PMID: 31308319]

- Elliott PM, Anastasakis A, Borger MA, et al. 2014 ESC Guidelines on [9] diagnosis and management of hypertrophic cardiomyopathy: the Task Force for the Diagnosis and Management of Hypertrophic Cardiomyopathy of the European Society of Cardiology (ESC). Eur Heart J 2014; 35(39): 2733-79. [http://dx.doi.org/10.1093/eurheartj/ehu284] [PMID: 25173338]
- [10] Kampmann C, Wiethoff CM, Wenzel A, et al. Normal values of M mode echocardiographic measurements of more than 2000 healthy infants and children in central Europe. Heart 2000; 83(6): 667-72.

[http://dx.doi.org/10.1136/heart.83.6.667] [PMID: 10814626]

- [11] Nagueh SF, Bierig SM, Budoff MJ, et al. American Society of Echocardiography clinical recommendations for multimodality cardiovascular imaging of patients with hypertrophic cardiomyopathy: Endorsed by the American Society of Nuclear Cardiology, Society for Cardiovascular Magnetic Resonance, and Society of Cardiovascular Computed Tomography. J Am Soc Echocardiogr 2011; 24(5): 473-98. [http://dx.doi.org/10.1016/j.echo.2011.03.006] [PMID: 21514501]
- [12] Richards S, Aziz N, Bale S, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med Off J Am Coll Med Genet 2015; 17(5): 405-24.
  - [http://dx.doi.org/10.1038/gim.2015.30] [PMID: 25741868]
- [13] Schrödinger L, DeLano W. PyMOL. 2020. Available from: http://www.pymol.org/pymol
- [14] Van Driest SL, Vasile VC, Ommen SR, et al. Myosin binding protein C mutations and compound heterozygosity in hypertrophic cardiomyopathy. J Am Coll Cardiol 2004; 44(9): 1903-10. [http://dx.doi.org/10.1016/j.jacc.2004.07.045] [PMID: 15519027]
- [15] Chiou K-R, Chu C-T, Charng M-J. Detection of mutations in symptomatic patients with hypertrophic cardiomyopathy in Taiwan. J Cardiol 2015; 65(3): 250-6. [http://dx.doi.org/10.1016/j.jjcc.2014.05.010] [PMID: 25086479]
- Jääskeläinen P, Vangipurapu J, Raivo J, et al. Genetic basis and outcome in a nationwide study of Finnish patients with hypertrophic [16] cardiomyopathy. ESC Heart Fail 2019; 6(2): 436-45. [http://dx.doi.org/10.1002/ehf2.12420] [PMID: 30775854]
- [17] Lopes LR. Brito D. Belo A. Cardim N. Genetic characterization and genotype-phenotype associations in a large cohort of patients with hypertrophic cardiomyopathy - An ancillary study of the Portuguese registry of hypertrophic cardiomyopathy. Int J Cardiol 2019; 278: 173-9

[http://dx.doi.org/10.1016/j.ijcard.2018.12.012] [PMID: 30554928]

- Otsuka H, Arimura T, Abe T, et al. Prevalence and distribution of [18] sarcomeric gene mutations in Japanese patients with familial hypertrophic cardiomyopathy. Circ J 2012; 76(2): 453-61. [http://dx.doi.org/10.1253/circj.CJ-11-0876] [PMID: 22112859]
- [19] Fujita E, Nakanishi T, Nishizawa T, Hagiwara N, Matsuoka R. Mutations in the cardiac troponin T gene show various prognoses in Japanese patients with hypertrophic cardiomyopathy. Heart Vessels 2013; 28(6): 785-94.

[http://dx.doi.org/10.1007/s00380-013-0332-3] [PMID: 23494605] [20] Thierfelder L, Watkins H, MacRae C, et al. a-tropomyosin and cardiac

- troponin T mutations cause familial hypertrophic cardiomyopathy: a disease of the sarcomere. Cell 1994; 77(5): 701-12. [http://dx.doi.org/10.1016/0092-8674(94)90054-X] [PMID: 8205619]
- Becker JR, Deo RC, Werdich AA, Panàkovà D, Coy S, MacRae CA. [21] Human cardiomyopathy mutations induce myocyte hyperplasia and activate hypertrophic pathways during cardiogenesis in zebrafish. Dis Model Mech 2011; 4(3): 400-10. [http://dx.doi.org/10.1242/dmm.006148] [PMID: 21245263]

## © 2022 Pham et al.

This is an open access article distributed under the terms of the Creative Commons Attribution 4.0 International Public License (CC-BY 4.0), a copy of which is available at: https://creativecommons.org/licenses/by/4.0/legalcode. This license permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.